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Structure and assembly cues for rhizospheric *nirK*- and *nirS*-type denitrifier communities in long-term fertilized soils



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ABSTRACT

The stimulatory effect of plants on soil denitrification activity has been widely reported, and root-derived carbon (C) is considered a factor contributing to enhanced denitrification activity in the rhizosphere. However, the mechanisms through which root-derived C shapes the rhizospheric denitrifying community structure remains unclear, especially under different soil fertility levels. Here, DNA-based stable isotope probing (DNA-SIP) and Illumina MiSeq sequencing were employed to characterize root-associated denitrifier communities containing nitrite reductase genes (nirS and nirK) in the rhizosphere of wheat grown in soils with three distinct long-term (32-year) fertilization regimes. Fertilization showed a significant impact on the composition of denitrifying communities that actively utilized photosynthetically fixed C in the wheat rhizosphere. In soils amended with inorganic fertilizers, wheat root-derived ¹³C was mainly assimilated by *nirS*-type denitrifiers affiliated to Alcaligenaceae and nirK-type denitrifiers affiliated to Phyllobacteriaceae. In contrast, organic fertilization resulted in larger diversity of ¹³C-labeled denitrifier communities where *nirS*-type denitrifiers such as Rhodobacteraceae and unclassified Burkholderiales and some unknown nirK-type denitrifiers were more abundant. The nirS-type denitrifier community was found to be more sensitive to the rhizosphere effect than the nirK-type community. Approximately 31% of the ¹³C-labeled nirS-type denitrifiers were more abundant in the rhizosphere than in the bulk soil, but only 2% of the ¹³C-labeled nirK-type denitrifiers showed increased abundance in the rhizosphere. The results of this study present direct evidence that root exudates can act as inducible C sources for heterotrophic denitrifying bacteria, but this induction pattern differs between nirS- and nirK-type communities and is dependent on soil fertility level.

1. Introduction

Nitrous oxide (N₂O) is a major stratospheric ozone-depleting substance and one of the most important greenhouse gases on Earth (Prentice et al., 2012; Blunden et al., 2013). The atmospheric concentration of N₂O has been increasing at nearly 0.75 ppb year⁻¹ since 1970 (IPCC, 2014). The increasing N₂O levels in the atmosphere are significantly associated with high nitrogen (N) and manure application rates in agricultural ecosystems (Syakila and Kroeze, 2011), and soil microbial denitrification plays a key role in this process. Denitrifying bacteria reduce soluble nitrate (NO₃⁻) or nitrite (NO₂⁻) to dinitrogen (N₂) through a series of gaseous intermediates, including nitric oxide (NO) and N₂O. Among these, the key reaction is reduction of NO₂⁻ to NO, which is catalyzed by two structurally different, but functionally equivalent NO₂⁻ reductases; namely, cytochrome cd1-containing reductase (NirS) and copper-containing reductase (NirK) (Hochstein and Tomlinson, 1988; Cutruzzola et al., 2001). This step causes dissolved N to become gaseous N for the first time during the denitrification process. Therefore, *nirS* and *nirK* genes are usually used as molecular markers to investigate the ecological behavior of denitrifying microorganisms in the environment. These two genes are thought to be mutually exclusive among denitrifying species and to represent two ecologically distinct denitrifying groups (Jones and Hallin, 2010). Several studies have shown that the responses of the *nirS*-type denitrifier community to environmental gradients are markedly different from those of the *nirK*-type denitrifier community, which supports the possibility that the two communities occupy different ecological niches (Jones and Hallin, 2010; Wei et al., 2015; Azziz et al., 2017).

Terrestrial plants release an array of substrates (i.e. rhizodeposition) that have the potential to lead to interactions between plants and

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